

**Resolution Number: AC/II(23-24).2.RUS7**

**S. P. Mandali's**  
**Ramnarain Ruia Autonomous College**  
*(Affiliated to Mumbai University)*



**Syllabus for UG**  
**Program: S.Y.B.Sc. (Minor) Life Science**

**Program Code: RUSLSc**

(As per the guidelines of National Education Policy 2020

For **Academic year 2024-25**)

(Choice based Credit System)

**Course Code: RUSLSc Minor I**  
**Course Title: Minor SY Sem III**  
**Academic year 2024-25**

SY Sem III Minor		<u>Minor</u>  <b>Biochemical Approach to Life Processes in Plants and Animals-I</b>	<b>3 Credits Total 45 lectures</b>
	<b>I</b>	<p><b>Enzymology</b></p> <p>Strategies for Isolation and purification of enzymes, measurement of specific activity and purification fold. Classification of enzymes (With an example of each). Effect of pH and Temperature.</p> <p>Coenzymes and cofactors : NAD, FAD, Mn, Mg, Zn and Cu (one example each).</p> <p>Kinetics (Michealis Menten, Lineweaver Burk plots). Enzyme Inhibitors, Activators and feed-back inhibition. Allosteric enzymes (Kinases in Glycolysis) and their significance in metabolic regulation.</p> <p>Concept of Isoenzymes: LDH.</p>	<b>15 Lectures</b>
	<b>II</b>	<p><b>Carbohydrate Metabolism and Bioenergetics.</b></p> <p>Carbohydrate Metabolism:</p> <p>a) Glycolysis – Process and metabolic regulation</p> <p>b) Pentose Phosphate Pathway</p> <p>c) Citric Acid Cycle: Process and regulation, Importance as a central amphibolic pathway.</p> <p>Electron Transport System: Localisation and Sequence of electron transporters.</p> <p>Oxidative Phosphorylation: Mitchell’s Chemiosmotic Hypothesis, ATP synthesis, Control of respiration, uncoupling and metabolic poisons.</p>	<b>15 Lectures</b>
	<b>III</b>	<p><b>Lipids and Proteins Metabolism.</b></p> <p>Lipids - Catabolism : Lipolysis, Role of Carnitine in mitochondrial permeability, Beta– Oxidation of fatty acids and integration into Kreb’s cycle, Ketone bodies and their significance.</p> <p>Amino Acids – Catabolism: Protein Degradation liberating amino-acids’, Deamination,</p>	<b>15 Lectures</b>

		Transamination & ammonia disposal by Urea cycle, Decarboxylation & integration into Krebs's cycle.	
		<b>PRACTICALS</b>	<b>1 Credit/ Total 15 lectures</b>
<b>RUSLScP MINOR</b>	<b>II</b>	<p><b>Practicals in Biochemical Approach to Life Processes in Plants and Animals- I</b></p> <p>1.A. Instrumentation / Technique</p> <ul style="list-style-type: none"> <li>- pH metry</li> <li>- Colorimetry</li> <li>- Titration.</li> </ul> <p>B- Process / Concept and immediate Relevance.</p> <ul style="list-style-type: none"> <li>- Extraction, Purification</li> <li>- Analysis / Estimation</li> <li>- GLP(Good Laboratory practices) incorporated into every practical</li> </ul> <p>Acid, bases and buffers.</p> <p>2. pH meter -</p> <ul style="list-style-type: none"> <li>a) Principle &amp; instrumentation and</li> <li>b) Determination of pH (titration of Acids/Bases/Buffers/ 'chameleon balls'). <i>(in FY the students were introduced to the concept of pH measurement of familiar liquids- here tech &amp; details are given- practically understanding buffering using Glycine / titration curve).</i></li> <li>c) Phosphate buffer preparation using Henderson Hasselbalch equation</li> <li>d) Glycine titration</li> </ul> <p>3. Protein precipitation by pH manipulation (Casein from Milk/ Curds) <i>(From previous experiments and pH manipulation, proteins can be precipitated).</i></p> <p>4. Study of Enzyme activity and Kinetics: Determination of <math>K_M</math> of an enzyme. Urease (from Jack beans) /Lipase/Protease/ (from seeds/ detergents) / amylase source <i>(Enzyme activity can be detected and estimated - using colorimetry).</i></p> <p>5. Histochemical localization of Enzymes (Acid Phosphatase) <i>(Enzyme activity can be localized).</i></p> <p>6. Estimation / Quantitation :</p>	

		<p>Colorimetric Protein Estimation by Biuret Method. (Enzyme extract / Casein from previous expts)  <i>(Proteins, such as the isolate from experiment 2 can be estimated by colour reaction).</i></p> <p>7. Colorimetric Cholesterol Estimation / total Lipid Estimation from egg. <i>(Lipid metabolism is an important component of our systems, content can be estimated by colour reaction).</i></p> <p>8. Titrimetric estimation of Ascorbic acid (Vit C). <i>(Estimation of biological materials by non-colorimetric method)</i></p>	
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## REFERENCES

	<b>RUSLSc 302</b>
1	1. Lehninger's Principles of Biochemistry Eds : D.L Nelson and M.M. Cox, Pub : WH Freeman Publishers, New York. 4th edition (2005)
2	Biochemistry Eds : J.M. Berg, J L Tymencko and L. Stryer  Pub : W H Freeman and co., New York. 5th edition (2002)
3	Fundamentals of Biochemistry by Eds : D.Voet , J. G. Voet Pub : John Wiley &Co., New York Pratt 1st ed (2004)
4	Principles of Biochemistry Ed: Lehninger.A Pub: CBS Publishers and Distributors, 2nd Edition (1993)
5	Principles of Biochemistry  Eds: Zubay G.L, Parson W.W. and Vance D.E. Pub: W. C. Brown, First Edition (1995)
6	An Introduction to Genetic Analysis Ed: Griffiths A.J. et al, Pub: W. H. Freeman London) Seventh Edition(2000)

## Modality of Assessment

### Theory Examination Pattern:

#### A) Internal Assessment- 40%- 40 Marks

Sr No	Evaluation Type	Marks
1.	Written Test	20
2.	Presentation on topic from syllabus / Quiz / Open book test	10
3.	Presentation on any journal article/ newsletter/ book review/ conference/ guest lecture	10
	<b>TOTAL</b>	<b>40</b>

#### B) External Examination- 60%- 60 Marks

### Semester End Theory Examination: (Deviation from the usual modality)

#### Semester End Theory Examination:

Duration - These examinations shall be of **2 HRS** duration.

Theory question paper pattern:

#### Paper Pattern:

Question	Options	Marks	Based on
Q1	Answer any 2 questions out of 3	16	Unit I
Q2	Answer any 2 questions out of 3	16	Unit II
Q3	Answer any 2 questions out of 3	16	Unit III
Q4	Short notes on topics of all 3 units; Answer any 3 out of 5;	12	Unit I, II, III
		<b>Total</b>	<b>60</b>

### Practical Examination Pattern:

#### A) Internal Examination:20 Marks

Particulars	Marks
Journal	05
Experimental tasks	15
<b>Total</b>	<b>20</b>

#### B) External Examination:30 Marks

##### Semester End Practical Examination:

Particulars	Paper
Main question to perform Experimental task/Estimation/ dissection/Bioinformatics statistical analysis/ project work	20
Identifications	10
<b>Total</b>	<b>30</b>

### Overall Examination & Marks Distribution Pattern

#### Semester III

Course	MINOR II			Grand Total
	Internal	External	Total	
Theory	40	60	100	200
Practical	20	30	50	100

**Course Code: RUSLSc Minor II**

**Course Title: Biochemical Approach to Life Processes in Plants and Animals-II  
(Minor SY Sem IV)**

**Academic year 2024-25**

**COURSE OUTCOMES:**

<b>COURSE OUTCOME</b>	<b>DESCRIPTION</b>
	<b>Students will gain insights about following;</b>
<b>CO 1</b>	To explain the anabolic pathways in carbohydrate, lipids and proteins
<b>CO 2</b>	To compare non- cyclic and cyclic photosynthetic pathways. To justify how photorespiration is a wasteful process and to review how C4 plants circumvent it.
<b>CO 3</b>	Understand the role of different enzymes in replication of DNA. Compare between the prokaryotic and eukaryotic replication process.
<b>CO 4</b>	Explain the transcription and translation process in prokaryotes and regulation of gene expression in prokaryotes.
<b>CO 5</b>	Compare transcription and translation process in eukaryotes. Understand the concept of post translational modifications in eukaryotes

<b>Minor - Sem IV</b>		<b>Minor - Biochemical Approach to Life Processes in Plants and Animals- II</b>	<b>3 Credits Total 45 lectures</b>
	<b>I</b>	<b>Anabolism of Biomolecules</b>  Anabolism of Carbohydrates: a) Gluconeogenesis b) Glycogen synthesis Anabolism of Lipids: a) Fatty acid biosynthesis b) Cholesterol and prostaglandin biosynthesis. Anabolism of Amino acids: a) Transamination and its significance b) Glutamine synthesis Synthesis of purines & pyrimidines with Significance. Photosynthesis, Light reaction and Calvin cycle Photorespiration in plants: C3 and C4 plants	<b>15 Lectures</b>
	<b>II</b>	<b>Molecular Biology studies in prokaryotes</b> DNA replication in prokaryotes. Transcription in Prokaryotes Translation in prokaryotes Regulation of gene expression and its significance: Operon model (Lactose / Tryptophan)	<b>15 Lectures</b>
	<b>III</b>	<b>Molecular Biology studies in eukaryotes</b> DNA Replication in Eukaryotes Transcription in Eukaryotes and Post Transcriptional modifications Translation in Eukaryotes and post translational modification	<b>15 Lectures</b>
<b>PRACTI CALS</b>		<b>PRACTICALS</b>	<b>1 Credit/ Total 15 lectures</b>
<b>RULScP MINOR SEM IV</b>		Practicals in Biochemical Approach to Life Processes in Plants and Animals- II. : A. Instrumentation / Technique ( I / T ) (1) PAGE (Demonstration). Chromatography – Paper, Thin layer, Column. B. Process / Concept and immediate Relevance (C and R ) - Extraction, Purification - Analysis / Estimation GLP(Good Laboratory practices) incorporated	



	<p>into every practical Separation / Extraction techniques</p> <p>1.Extraction and Detection of RNA/Ribose Sugars. C, T (<i>Extraction of nucleic acid and detection by colour reaction</i>)</p> <p>2.Chromatography of Sugars – Circular Paper C, T (<i>Separation of carbohydrates and detection by colour reaction</i>)</p> <p>3.Thin Layer Chromatography for separation of Plant Pigments.(Slide technique) C,T,R (<i>Separation techniques for charged, uncharged materials based on solvent partition</i>)</p> <p>4.Solvent Extraction of Lipids. C, T, R (<i>Extraction of lipid and proportional estimation by weight</i>)</p> <p>5.Column Chromatography of Proteins / Pigments. I, C, T(<i>Separation technique for proteins/ other materials based on charge/size</i>)</p> <p>6.Protein separation by PAGE (Demonstration) I, C (<i>Separation techniques for charged materials based on electrophoretic mobility</i>)</p> <p>7.Interpretation of pathological reports based on biochemical analysis.</p>	
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## REFERENCES

	<b>RUSLSc 402</b>
1	1. Lehninger's Principles of Biochemistry Eds : D.L Nelson and M.M. Cox, Pub : WH Freeman Publishers, New York. 4th edition (2005)
2	Biochemistry Eds : J.M. Berg, J L Tymencko and L. Stryer  Pub : W H Freeman and co., New York. 5th edition (2002)
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<b>Experimental tasks</b>	15
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**D) External Examination:30 Marks**  
**Semester End Practical Examination:**

<b>Particulars</b>	<b>Paper</b>
<b>Main question to perform Experimental task/Estimation/ dissection/Bioinformatics statistical analysis project work</b>	20
<b>Identifications</b>	10
<b>Total</b>	<b>30</b>

**Overall Examination & Marks Distribution Pattern**

**Semester IV**

<b>Course</b>	<b>MINOR II</b>		<b>Total</b>	<b>Grand Total</b>
	<b>Internal</b>	<b>External</b>		
<b>Theory</b>	<b>40</b>	<b>60</b>	<b>100</b>	<b>200</b>
<b>Practical</b>	<b>20</b>	<b>30</b>	<b>50</b>	<b>100</b>

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